

Isoforms of serum γ -glutamyltransferase in epileptic patients treated with enzyme-inducing anticonvulsant drugs.

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ABSTRACT PURPOSE. The increase of serum activity of γ -glutamyltransferase (γ GT) through the action of enzyme-inducing anticonvulsant drugs has been widely documented; however, the behaviour of its multiple forms and its relationship with the degree of enzyme induction has received little coverage. This subject is the major aim of our paper. **METHODS.** An electrophoretic study of the serum γ GT isoforms was made in 90 adult epileptic patients under chronic treatment with phenobarbital, phenytoin and carbamazepine in polytherapy. **RESULTS.** A significant correlation was found ($p < 0.001$) between the drug score and urinary excretion of D-glucaric acid (DGA) ($r = 0.773$), total γ GT ($r = 0.382$), γ GT1 ($r = 0.398$) and γ GT2 ($r = 0.361$). In a group of 11 patients with the γ GT3 isoforms, considered a sensitive test for cholestasis, serum activities of total γ GT, γ GT1, γ GT2 and 5'-nucleotidase were found that were significantly higher than in the 79 patients without the γ GT3 isoform ($p < 0.001$); however, for the drug score and excretion of DGA, no significant differences were found, suggesting a similar degree of enzyme induction in both groups of patients. **CONCLUSIONS.** The presence of cholestasis, at least biochemically detectable in some of these patients, appears to be a factor of paramount importance when interpreting the effect of enzyme-inducing anticonvulsant drugs on serum γ GT. This fact may contribute towards explaining its highly varied response to the administration of these drugs.

INTRODUCTION

Enzyme induction by drugs mostly concerns the enzyme systems involved in drug metabolism. γ -Glutamyltransferase (γ GT, EC 2.3.2.2) is a membrane-bound enzyme that participates in the metabolism of glutathione, cleaving its γ -glutamyl peptide bond and

transferring the glutamyl moiety to acceptor molecules. As a result, and via glutathione, γ GT is indirectly involved in drug metabolism (1).

Hepatic γ GT is induced by phenobarbital-type enzyme-inducing agents, and increased serum enzyme activities have been described in patients treated with anticonvulsant drugs by different authors (2-4). However, as well as the induction of the enzyme protein synthesis in the liver, other mechanisms such as alterations in the lipid composition of plasma membranes may contribute to the increase of γ GT serum activity in these patients (5,6). γ GT has a well-documented enzyme heterogeneity (7), and its serum multiple forms have been previously studied in patients treated with anticonvulsant drugs (8-10), although this subject has been dealt with in a horizontal manner. The urinary excretion of D-glucaric acid (DGA), an end product of carbohydrate metabolism in humans produced via the glucuronic acid pathway has been widely used as an indirect enzyme induction marker and it is significantly increased by these drugs (1).

In our article, we present the results obtained for the isoforms of serum γ GT in a group of adult epileptic patients treated with phenobarbital, phenytoin and carbamazepine, in an attempt to clarify its relationship with the degree of enzyme induction evaluated by means the drug score and the urinary excretion DGA.

MATERIAL AND METHODS

A group of 90 epileptic patients (56 males and 34 female) with a mean age (\pm SEM) of 38.0 ± 1.5 years was studied, who had been treated for more than 10 years with phenobarbital ($n = 60$), phenytoin ($n = 70$) and carbamazepine ($n = 33$). In all cases, there was adequate therapeutic compliance, and no additional pharmacological treatment was received. As the anticonvulsant drugs were administered in polytherapy, the dose was expressed as units/day, according to a drug score in which one unit corresponded to every

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30 mg of phenobarbital, 50 mg of phenytoin and 100 mg of carbamazepine (2, 11). After an overnight fast, venous blood and urine samples were taken before the morning administration of anticonvulsant drugs, whose doses had not been modified for at least three months beforehand. The control group comprised 49 medication-free clinically healthy individuals (30 male and 19 female) with a mean age of 36.7 ± 1.6 years. Pregnant women or those who were taking oral contraceptives were excluded from both the control and patients group.

The serum enzyme activities of γ GT were determined using γ -glutamyl-3-carboxy-4-nitroanilide as substrate using commercial reagents from Roche Diagnostics. The residual enzymatic activity after treatment of the serum samples with butanol was determined according to a previously described procedure (12, 13). Electrophoretic separation of the γ GT multiple forms was carried out on cellulose acetate plates (8). Serum activity of alcohol dehydrogenase (ADH, EC 1.1.1.1) was determined spectrophotometrically (14), and serum α -glutathione-S-transferase (α GST, EC 2.5.1.18) was determined using an enzyme immunoassay commercialized by Biotrin International. The serum activities of aspartate aminotransferase (AST, EC 2.6.1.1) and alanine aminotransferase (ALT, EC 2.6.1.2) were determined according to the recommendations of the Spanish Clinical Biochemistry Society. The activity of 5'-nucleotidase (5'NU, EC 3.1.3.5) was determined using commercial reagents from Sigma Diagnostics. Urinary DGA was determined using an enzymatic procedure (15), and the results were expressed as the ratio of DGA to creatinine urinary concentrations (16). The variations of the different variables studied in the patient group were calculated using the expression: Variation (%) = 100 (median patients - median controls) / median controls.

Statistical analysis of the data was carried out using Microsoft Excel (v.5.0). The Kolmogorov-Smirnov test was applied to check for normality. Parametric tests were used when the data had a Gaussian distribution (Student's t test and Pearson's correlation coefficient); otherwise, non-parametric tests were used (Mann-Whitney test and Spearman's correlation coefficient). The results were expressed as mean \pm SEM (median).

RESULTS

The urinary excretion of DGA in the patient group was 276.4 ± 22.8 (244.5) μ mol/g creatinine, and in the control group 18.5 ± 1.0 (16.7) μ mol/g creatinine, with a highly significant difference between both groups ($p < 0.001$). In 93% of the patients, urinary DGA was higher than the upper reference limit.

For serum γ GT, the patient group presented a significantly higher activity than the control group (Table 1), with 81% of the cases presenting activities higher than the corresponding upper reference limit for their sex.

Table 1: Serum enzyme activities in the control and patient groups.

| | Controls (n=49) | Patients (n=90) | Change (%) |
|---------------------------|-----------------------|--------------------------------------|------------|
| Total γ GT (U/L) | 16.9 \pm 1.3 (14.0) | 76.3 \pm 7.2 (58.8) ^{***} | 320.0 |
| γ GT1 (U/L) | 5.2 \pm 0.7 (3.5) | 48.5 \pm 6.1 (27.5) ^{***} | 685.7 |
| γ GT2 (U/L) | 13.1 \pm 1.1 (12.1) | 24.6 \pm 1.3 (22.0) ^{***} | 81.8 |
| γ GT3 (U/L) | 0.0 \pm 0.0 (0.0) | 2.2 \pm 0.7 (0.0) | 0.0 |
| Residual γ GT (%) | 92.3 \pm 1.6 (100) | 78.2 \pm 1.3 (78.0) ^{***} | -22.0 |
| 5'NU (U/L) | 5.9 \pm 0.2 (6.0) | 8.3 \pm 0.2 (8.0) ^{***} | 33.3 |
| AST (U/L) | 25.1 \pm 1.3 (22.5) | 27.4 \pm 0.8 (25.0) [*] | 11.1 |
| ALT (U/L) | 25.2 \pm 2.1 (20.5) | 29.7 \pm 1.5 (26.0) ^{**} | 26.8 |
| ADH (U/L) | 0.13 \pm 0.01 (0.1) | 0.25 \pm 0.02 (0.2) ^{***} | 81.8 |
| α GST (μ g/L) | 2.03 \pm 0.21 (1.7) | 4.29 \pm 0.62 (3.0) ^{***} | 76.5 |

Significance: * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$

Using the electrophoretic technique applied, two isoforms of serum γ GT were separated in the control subjects, one with α 1-globulin mobility (γ GT1) and another with α 2-globulin mobility (γ GT2) with higher relative activity (see Table 1). In the patient group, a significant increase ($p < 0.001$) was found for total γ GT, γ GT1 and γ GT2 activity, with a significant increase ($p < 0.001$) in the relative proportion of γ GT1 as compared to the control group ($53.9 \pm 2.2\%$ vs. $27.0 \pm 2.5\%$). In 11 of the patients studied (12.2%) an additional isoform was found with β -globulin mobility (γ GT3) that is considered as a sensitive test for cholestasis (17, 18). Amongst the patients studied, the total γ GT activities had a significant correlation with the activities of the γ GT1 isoform ($r = 0.963$, $p < 0.001$) and γ GT2 isoform ($r = 0.630$, $p < 0.001$), as well as with the relative proportion as a percentage of γ GT1 ($r = 0.765$, $p < 0.001$). These results show that the increases in serum γ GT activity in these patients is mainly due to the γ GT1 isoform.

Table 1 shows the results obtained for γ GT and its multiple forms, as well as for the other biochemical variables studied, in the control and patient groups. With the exception of the activity of the γ GT3 isoform, significant differences were found between both groups for all of the variables, although their variations with relation to the control group are highly varied. The residual γ GT activity after treatment with butanol was significantly higher in the control group ($p < 0.001$), which suggests that the γ GT1 isoform is affected more than γ GT2. This was confirmed by electrophoretically fractioning the γ GT isoforms in serum samples from patients before and after treatment with butanol. Moreover, in the patient group a significant negative correlation was found between the residual γ GT activity and the relative proportion as a percentage of the γ GT1 isoform ($r = -0.559$, $p < 0.001$).

A highly significant correlation was found in the patient group between the urinary excretion of DGA and the drug score ($r = 0.773$, $p < 0.001$). Table 2 shows the correlation coefficients for γ GT and its multiple forms with the drug score, urinary DGA and serum 5'NU activity. Significant correlations were also found between γ GT and AST, ALT, ADH and α GST ($p < 0.005$).

Table 2: Relationship between serum γ GT isoforms and other variables in the patient group.

| | Total γ GT | γ GT1 | γ GT2 |
|------------|-------------------|--------------|--------------|
| Drug score | 0.382** | 0.398** | 0.361** |
| DGA | 0.380** | 0.384** | 0.329* |
| 5'NU | 0.587** | 0.583** | 0.340* |

Significance: * $p < 0.005$; ** $p < 0.001$

Table 3 shows the results obtained from making a comparison of the patients studied depending on whether or not they had the γ GT3 isoform. Although no significant differences were found for the drug score and the urinary excretion of DGA, the serum activities of total γ GT, γ GT1, γ GT2 and 5'NU were significantly higher in the patients who presented the γ GT3 isoform; however, the increase was much higher for γ GT1 (388%) than for γ GT2 (86%). A dichotomy of the patients according to sex not appear to offer any information with additional interest.

Table 3: Serum activities of γ GT isoforms and other variables in the groups of patients without and with the γ GT3 isoform.

| | Without γ GT3 (n=79) | With γ GT3 (n=14) |
|-------------------------------|-----------------------------|----------------------------|
| Total γ GT (U/L) | 57.1 \pm 4.0 (51.0) | 214.2 \pm 27.1 (174.0)** |
| γ GT1 (U/L) | 34.5 \pm 3.8 (25.2) | 155.1 \pm 26.6 (123.0)** |
| γ GT2 (U/L) | 22.8 \pm 1.2 (20.9) | 41.1 \pm 3.4 (38.8)** |
| γ GT3 (U/L) | 0.0 \pm 0.0 (0.0) | 17.9 \pm 2.5 (19.0)** |
| Residual γ GT (%) | 79.7 \pm 1.4 (79.0) | 68.1 \pm 3.3 (72.0)* |
| 5'NU (U/L) | 7.9 \pm 0.2 (8.0) | 11.4 \pm 0.8 (11.0)** |
| Drug score (units/day) | 9.8 \pm 0.6 (8.9) | 9.3 \pm 1.0 (9.9) |
| DGA (μ mol/g creatinine) | 282.1 \pm 24.4 (244.0) | 235.6 \pm 15.4 (242.3) |

Significance: * $p < 0.01$; ** $p < 0.001$

DISCUSSION

Judging by the results obtained for the urinary excretion of DGA, the studied epileptic patients presented a high degree of enzyme induction. The degree of enzyme induction produced by anticonvulsant drugs is dose-dependent (19), and the excretion of DGA revealed a high correlation with the drug score used ($r = 0.765$, $p < 0.001$), which may reflect the enzyme-inducing capacity of the anticonvulsant drugs administered in polytherapy (11, 20).

Similarly, the patients studied presented a significant increase in serum γ GT activity (see Table 1). Treatment with phenobarbital-type enzyme-inducing agents produced a strong induction of γ GT in the hepatic plasma membranes (21); however, enzyme induction is not the only mechanism that may be involved in the increase of γ GT serum activity (1). Another important factor is the possible effect of these drugs on the lipid composition of plasma membranes, by increasing the phospholipids/cholesterol molar ratio (5, 6). This ratio is the most determining factor for the fluidity of the membranes, as well as the interaction between different intramembrane composites, and as a result, the drug treatment may facilitate the solubilization and release of γ GT hydrophobic forms from the plasma membranes to the extracellular milieu (5, 6). In blood, the hydrophobic forms of γ GT may form aggregates with the lipoprotein X, and low (LDL) and very low density lipoproteins (VLDL), with an electrophoretic mobility similar to that of the β - γ globulins, whereas aggregates with high density lipoproteins (HDL) have α 1 mobility (22, 23). In clinically healthy individuals, the predominant γ GT forms are hydrophylic (24) and do not form complexes with the lipoproteins (22, 25), although they may have α 1 mobility (22).

The patients studied presented an increase in the relative proportion of the $\alpha 1$ mobility isoform (γ GT1), whose hydrophobic nature explains the lower residual enzymatic activity after treatment of serum samples with butanol as compared to the control group, in which enzymatic activity was practically unchanged (see Table 1). The anticonvulsant drugs may favour the solubilization of γ GT bound to the plasma membranes and their release to the blood stream, where its hydrophobic nature allows them to form complexes with HDL constituting the γ GT1. This isoform could correspond to the intermediate relative molecular fraction, and the γ GT2 isoform to the low relative molecular form described by other authors (7, 9). Although the membrane-bound 5'NU is not induced by phenobarbital (26), its serum activity is also significantly increased in the patient group, having a better correlation with the γ GT1 than with the γ GT2 isoform (table 2). The serum activity of γ GT1 increased greatly in the patients studied (686%), whereas the possibly hydrophilic form γ GT2 had a much more moderate increase (82%), similar to that of the cytosolic enzymes ADH (82%) and α GST (77%) which are considered sensitive markers for hepatocellular damage (14, 27). The alteration of the lipid composition and the permeability of plasma membranes by the administered anticonvulsant drugs may also favour the release of cytosolic enzymes into the bloodstream.

The increases found in the group of patients for the cytosolic/mitochondrial AST (11%) and cytosolic ALT (27%) serum activities were lower than those obtained for ADH and α GST. This may be due to the preferable localization of both aminotransferases in azinar zone 1 (periportal), whereas the hepatic injury produced by drugs is mainly localized in azinar zone 3 (centrilobular), where the concentration of drugs and their metabolites is higher (28, 29). In turn, ADH is preferably localized in the centrilobular region (30), and α GST is equally distributed throughout the liver lobe (31). α GST, one of a family of detoxication enzymes, may be induced by phenobarbital although with a large organ and species variability (32).

The increase of serum 5'NU is specific for cholestatic liver injury, as the detergent action of bile acids on the canalicular membrane is the only mechanism for enzyme release into plasma (29). In the group of patients with the γ GT3 isoform, which as previously

mentioned is a sensitive test for cholestasis (17, 18), significant increases in 5'NU were found, as well as for total γ GT, γ GT1 and γ GT2 ($p < 0.001$), as compared to the patients who did not have γ GT3 isoform (see Table 3). However, no significant differences were found for the drug score or urinary excretion of DGA, which suggests a similar degree of enzyme induction in both groups of patients. This γ GT3 isoform could correspond to the high relative molecular mass fraction formed by aggregates of γ GT with LDL, VLDL or membrane fragments (7, 9). The prevalence of 12% for the presence of γ GT3 isoform in our group of patients is clearly lower than that of 21% detected by Kok et al. in a group of 38 psychiatric patients treated with phenobarbital and phenytoin (8). Possibly, the patients studied by these authors were concomitantly treated with neuroleptic, antipsychotic or antidepressant drugs.

Different factors have been indicated that should be taken into account when interpreting γ GT serum activity as an index for enzyme induction in patients treated with anticonvulsant drugs (3). Our results indicate that the presence of cholestasis, at least biochemically detectable in some of these patients, is a factor of extreme importance in explaining the increase in serum γ GT through the action of anticonvulsant drugs. In a similar way to that described for the ingestion of alcohol (33), the degree of response of γ GT to the administration of these drugs appears to be highly variable, which would explain the discrete correlation found between its serum enzymatic activity and the drug score (see Table 2). Similarly, in a previous study we found that in patients treated with anticonvulsant drugs there was no significant correlation between the changes in the serum activity of γ GT and those of drug score (20). As a result, the serum activity of total γ GT is of no use as a marker for enzyme induction, and separating the multiple forms of the enzyme does not appear to offer any additional information of practical interest in this sense. However, in epileptic patients, the evaluation of the γ GT3 isoform may be advisable in its biochemical monitoring as a sensitive marker of cholestasis produced by anticonvulsant drugs.

REFERENCES

- [1] Batt AM, Siest G, Magdalou J, Galteau MM. Enzyme induction by drugs and toxins. *Clin Chim Acta* 1992;209:109-121.

- [2] Rosalki SB. Plasma enzyme changes and their interpretation in patients receiving anticonvulsant and enzyme-inducing drugs. In: Richens A, Woodford FP, editors. *Anticonvulsant Drugs and Enzyme Induction*. Associated Scientific Publishers, Amsterdam 1976. p. 27-35.
- [3] Braide SA, Davies TJ. Factors that affect the induction of gamma glutamyltransferase in epileptic patients receiving anticonvulsant drugs. *Ann Clin Biochem* 1987;24:391-399.
- [4] Aldenhövel HG. The influence of long-term anticonvulsant therapy with diphenylhydantoin and carbamazepine on serum gamma-glutamyltransferase, aspartate aminotransferase, alanine aminotransferase and alkaline phosphatase. *Eur Arch Psychiatr Neurol Sci* 1988;237:312-316.
- [5] Ratanasavanh D, Magdolou J, Antoine B, Galteau MM, Siest G. Gamma glutamyltransferase activity of liver plasma membranes in phenobarbital-treated rabbits. *Pharmacol Res Commun* 1981;13:909-919.
- [6] Ratanasavanh D, Tazi A, Gaspart E, Jacquier A, Notter D, Galteau MM, Siest G. Hepatic gamma-glutamyltransferase release: Effect of bile salts and membrane structure modifications. In: Siest G, Eusghen C, editors. *Gamma-glutamyltransferases*. Advances in Biochemical Pharmacology. Masson, Paris 1982. p. 92-103.
- [7] Wenham PR, Horn DB, Smith AF. Physical properties of g-glutamyltransferase in human serum. *Clin Chim Acta* 1984;141:205-218.
- [8] Kok PJMJ, Seidel B, Holtkamp HC, Huisman J. A new procedure for the visualization of multiple forms of gamma-glutamyltransferase. Results in normals, patients receiving enzyme-inducing drugs and patients having liver parenchymal lesions. *Clin Chim Acta* 1978;90:209-216.
- [9] Wenham PR, Horn DB, Smith AF. Multiple forms of g-glutamyltransferase: A clinical study. *Clin Chem* 1985;31:569-573.
- [10] ellini M, Tumino E, Giordani R, Fabrini G, Costa F, Galli R, Rucco M, Belcari C, Michelassi C, Murri L, Maltinti G, Marchi S. Serum gamma-glutamyltransferase isoforms in alcoholic liver disease. *Alcohol Alcohol* 1997;32:259-266.
- [11] Richens A, Rowe DJF. Disturbance of calcium metabolism by anticonvulsant drugs. *Br Med J* 1970;4:73-76.
- [12] Beck PR. Butanol extraction of serum and urinary gamma-glutamyltransferase. In: Goldberg DM, Wilkinson SH, editors. *Enzymes in Health and Disease*. Karger, Basel 1978. p. 137-139.
- [13] Beck PR. Butanol extraction of serum and urinary g-glutamyltransferase and its application in clinical diagnosis. *Ann Clin Biochem* 1978;15:151-156.
- [14] Khayrollah AA, Al-Tamer YY, Taka M, Skursky L. Serum alcohol dehydrogenase activity in liver diseases. *Ann Clin Biochem* 1982;19:35-42.
- [15] Fernández MP, Tutor JC, Paz JM. Determinación del ácido D-glucárico urinario por un procedimiento enzimático. *Clin Chem Newsletter* 1982;2:77-81.
- [16] Makki KA, Beetham R, Richens A. Overnight urine specimens for the determination of D-glucuric acid excretion in man. *Br J Clin Pharmacol* 1979;8:183-186.
- [17] Nemesánszky E, Lott JA. Gamma-glutamyltransferase and its isoenzymes. Progress and problems. *Clin Chem* 1985;31:797-803.
- [18] Burlina A. Improved method for fractionating g-glutamyltransferase by electrophoresis on cellulose acetate. *Clin Chem* 1978;24:502-504.
- [19] Perucca E, Edges A, Makki KA, Ruprah M, Wilson JF, Richens A. A comparative study of the relative enzyme inducing properties of anticonvulsant drugs in epileptic patients. *Br J Clin Pharmacol* 1984;18:401-410.
- [20] Hermida J, Fernández MP, Tutor JC. Relationship between changes in drug score, D-glucuric acid excretion, and g-glutamyltransferase and b-glucuronidase serum activities during anticonvulsant treatment. *Clin Lab* 2002;48:115-119.
- [21] Ratanasavanh D, Tazi A, Galteau MM, Siest G. Localization of gamma glutamyltransferase in subcellular fractions of rat and rabbit liver: Effect of phenobarbital. *Biochem Pharmacol* 1979;28:1363-1365.
- [22] Saccheti L, Castaldo G, Salvatore F. Electrophoretic behaviour and partial characterization of disease-associated serum forms of gamma-glutamyltransferase. *Electrophoresis* 1989;10:619-627.
- [23] Huseby NE. Multiple forms of serum gamma-glutamyltransferase. Association with lipoproteins. *Clin Chim Acta* 1982;124:103-112.
- [24] Selvaraj P, Rolston DDK, Balasubramanian KA. Separation of hydrophobic and hydrophilic forms of g-glutamyltransferase from human serum by hydrophobic chromatography on phenylsepharose CL-4B. *Clin Chim Acta* 1984;138:141-149.
- [25] Artur Y, Wellman-Bednawska M, Jacquier A, Siest G. Complexes of serum gamma-glutamyltransferase with apolipoproteins and immunoglobulins. *Clin Chem* 1984;30:361-333.
- [26] Seifert J, Vácha J. Depression of microsomal 5'-nucleotidase and cellular ribonucleases in rat liver

- after phenobarbital administration. *Chem Biol Interactions* 1970;2:297-307.
- [27] Rees GW, Trull AK, Doyle S. Evaluation of an enzyme-immunometric assay for serum a-glutathione S-transferase. *Ann Clin Biochem* 1995;32:575-583.
- [28] Desmet VJ. Drug-induced liver disease: Pathogenic mechanisms and histopathological lesions. *Eur J Med* 1993;2:36-47.
- [29] Sturgill MG, Lambert GH. Xenobiotic-induced hepatotoxicity: mechanisms of liver injury and methods of monitoring hepatic function. *Clin Chem* 1997;43:1512-1526.
- [30] Buehler R, Hess M, Von Wartburg JP. Immunohistochemical localization of human liver alcohol dehydrogenase in liver tissue, cultured fibroblasts, and HeLa cells. *Am J Pathol* 1982;108:89-99.
- [31] Hayes PC, Bouchier IAD, Beckett GJ. Glutathione S-transferase in humans in health and disease. *Gut* 1991;32:813-818.
- [32] Ramana KV, Kohli KK. Differential effects of phenobarbitone on the hepatic and renal Glutathione-S-transferases in the rhesus monkey. *Indian J Pharmacol* 1998;30:34-37.
- [33] Matsuka Y, Wang DH, Suganuma N, Imai K, Ikeda S, Takeka K, Kira S. Differential responses of serum gamma-glutamyltransferase to alcohol intake in Japanese males. *Acta Med Okayama* 2003;57:171-78.